

Outline

- Scanning Probe Microscopy
- Atomic Force Microscopy
 - General set-up & operation modes
- Applications
 - Imaging mode
 - Force-distance mode
- Sample preparations
- Conclusion

Scanning probe microscopy (SPM)

~1600 Light Microscope

1938: Transmission Electron Microscope

1964: Scanning Electron Microscope

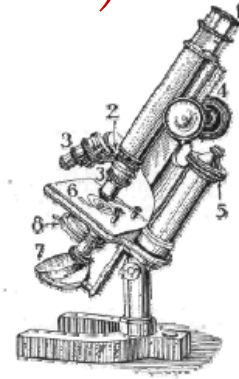
1982: Scanning Tunneling Microscope

1984: Scanning Near-field
Optical Microscope

1986: Atomic Force Microscope
- magnetic force, lateral force, chemical force...

When a conducting tip is brought very near to the surface to be examined, a bias (voltage difference) applied between the two can allow electrons to tunnel through the vacuum between them. The resulting *tunneling current* is a function of tip position, applied voltage, and the local density of states (LDOS) of the sample (Resolution: x-y: 0.1 nm)

Gerd Binnig and Heinrich Rohrer (1981) Nobel prize



Impression..

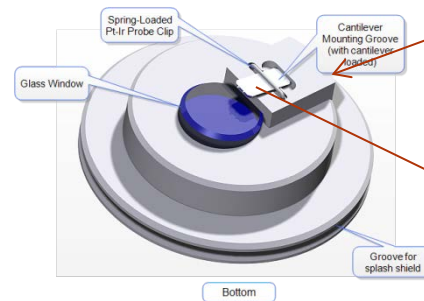


Typical resolution:
X-Y: 1 nm; Z: 0.1 nm

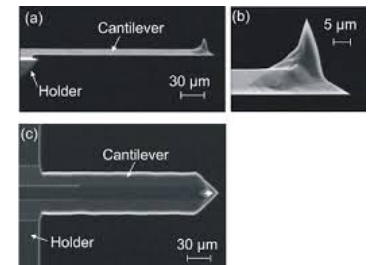
Using Dimension 3100 with Nanoscope IV controller (open-loop), possible scan size is $100 \times 100 \mu\text{m}^2$



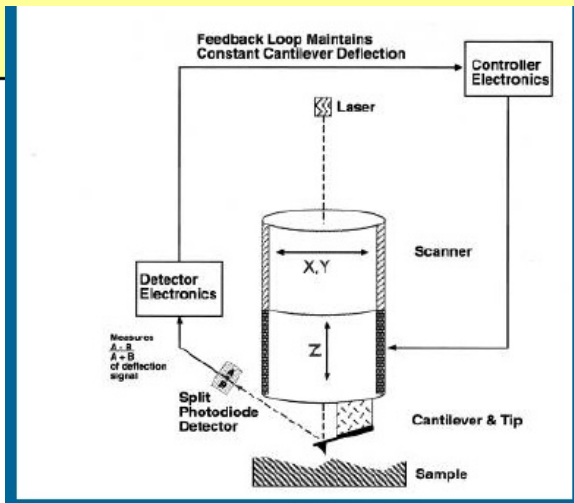
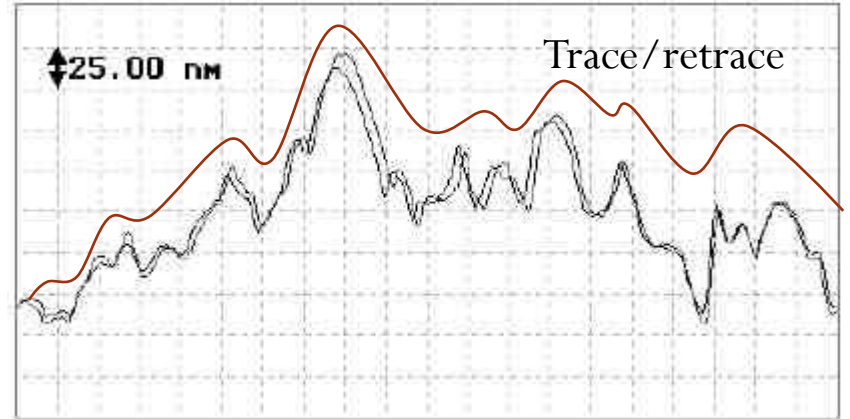
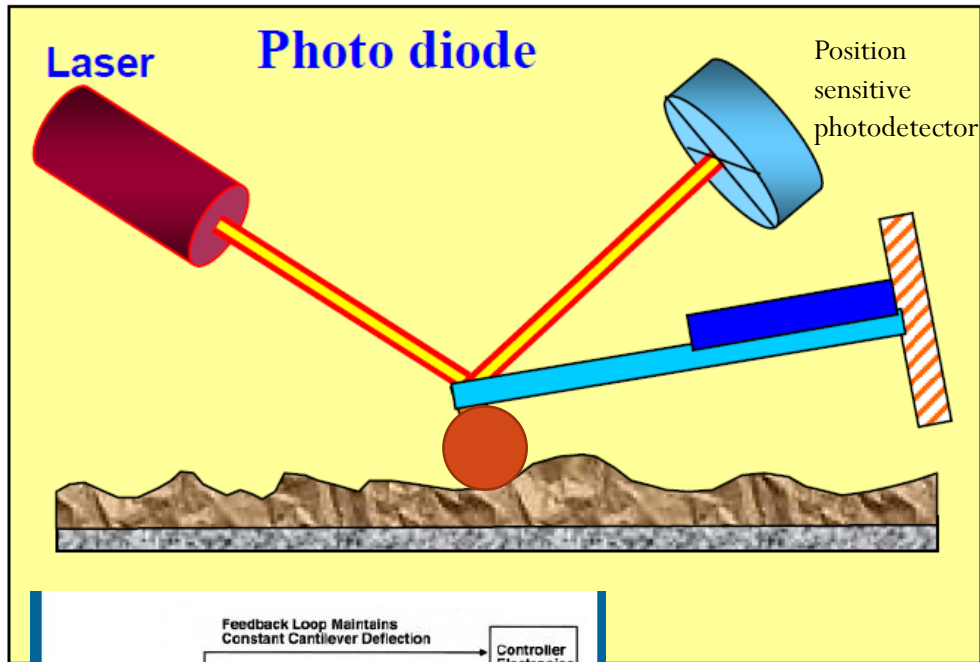
Scanner/
Piezo



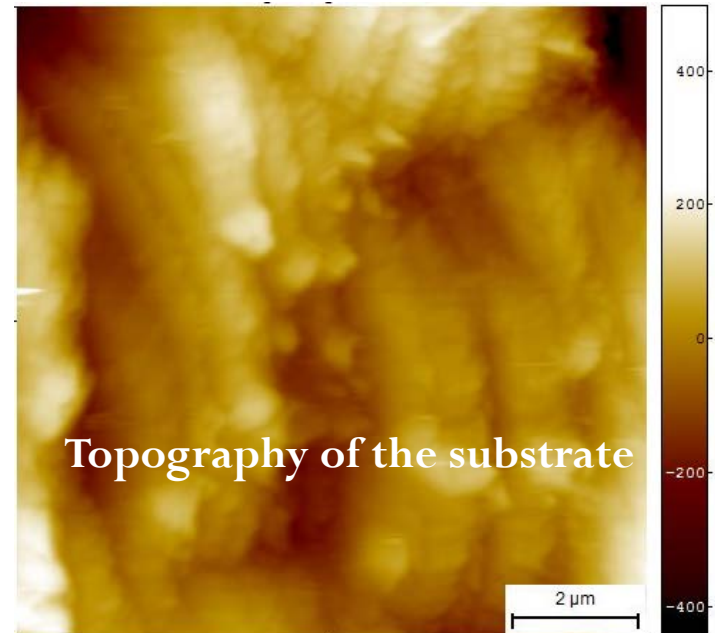
Cantilever holder



How AFM works?

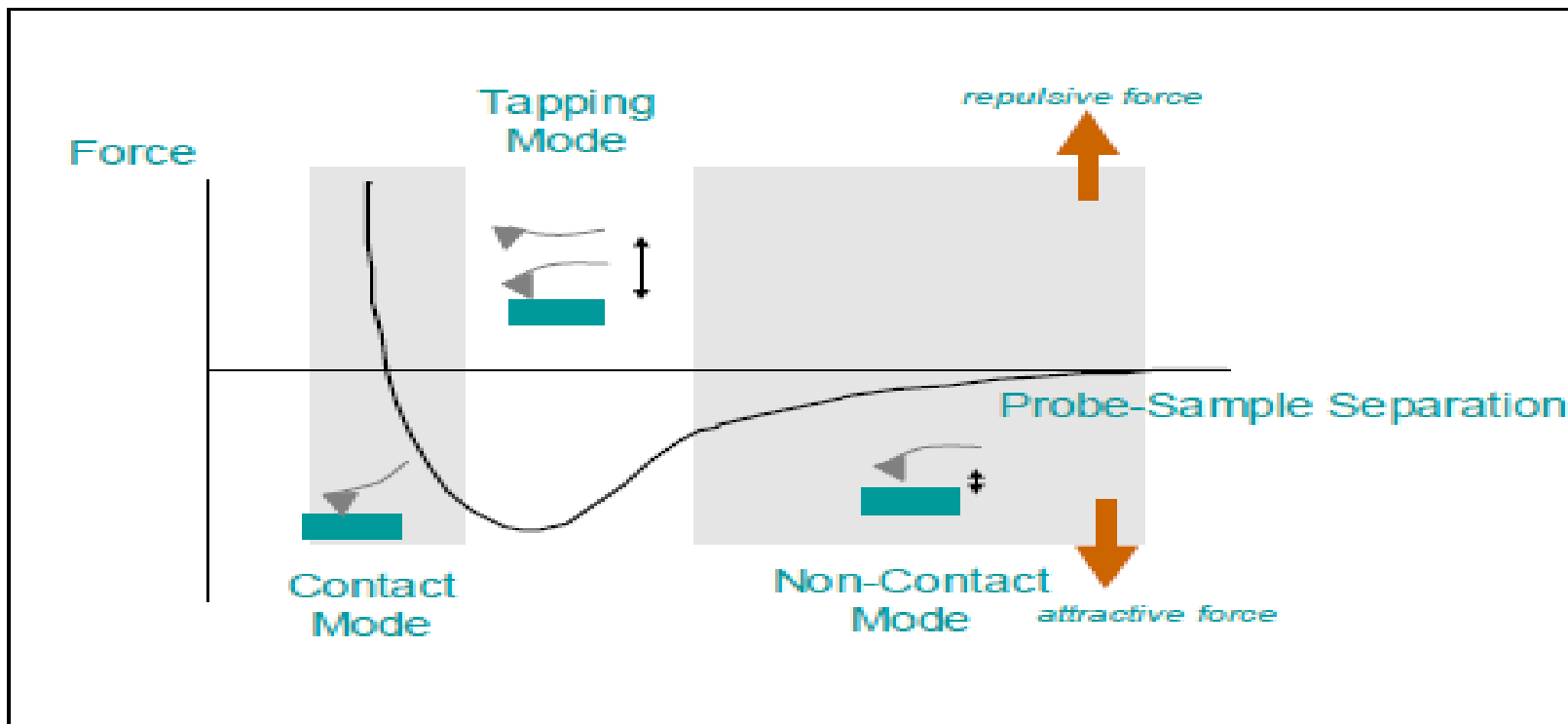


Hooke's law
 $F = -k \cdot x$



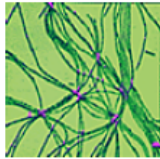
AFM operating modes

- Topographic modes:
 - **Contact mode** (Static mode) (<0.5 nm separation distance)
 - **Tapping/intermittent** mode (Amplitude modulation) (0,5-2 nm)
 - **Noncontact** mode (Frequency modulation) (0,1-10 nm)

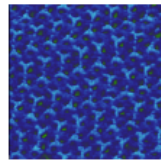


Applications of AFM

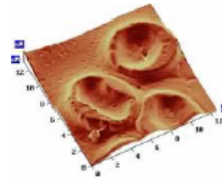
Lifescience



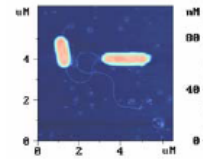
Actin filaments



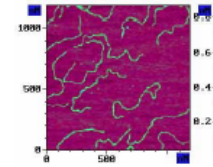
Proteins



Erythrocytes

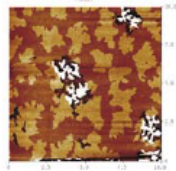


Bacteria

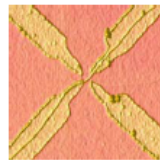


Linearised DNA

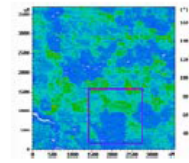
Materials and Surface Science



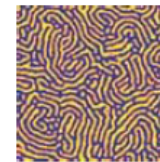
Organic film



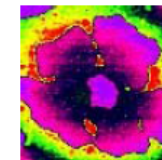
Transistor



Ferroelectric domains



Triblock copolymer film



Polymer

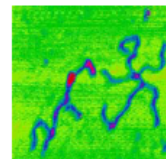
Nanolithography & Nanomanipulation



Polymer film engraving



Anodic oxidation

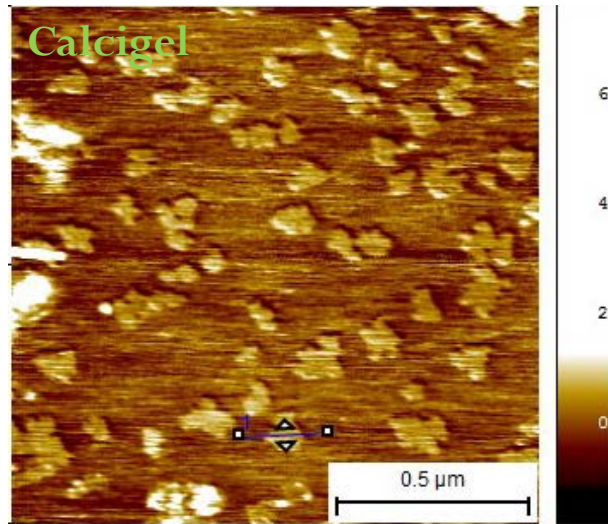


DNA on mica

Applications in our context

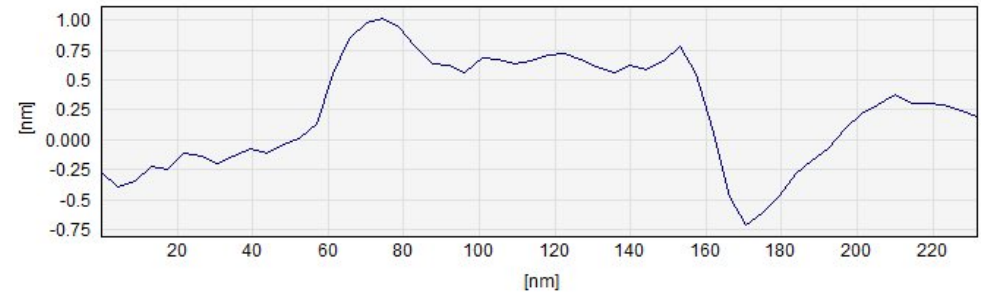
- Characterization of clay/nanoparticle size/morphology
- Topography of rock and mineral surfaces
 - Mineral dissolution, precipitation etc.
- Forces between surfaces using colloid probe technique
 - Mineral colloids
 - In the presence of electrolytic media

Clay particle characterization

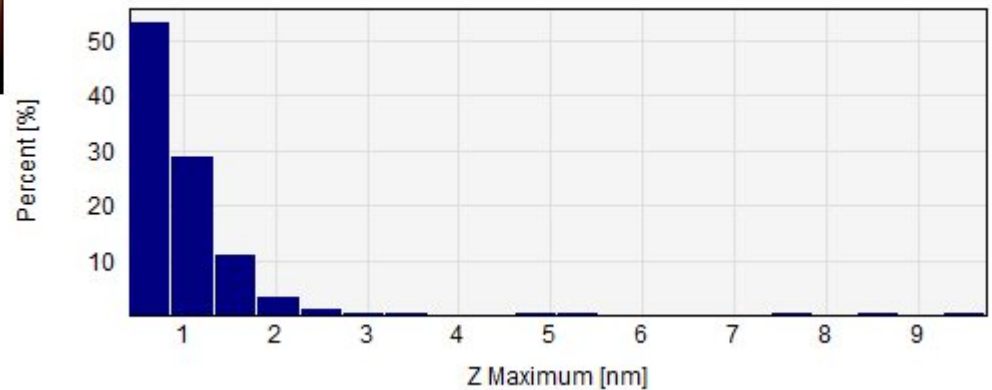


Topographic image

Parameters obtained:
Size, shape, height distribution



Height profile along the highlighted line



Particle height distribution histogram

Topography of rock and mineral surfaces

Image obtained with 1 μm colloid probe

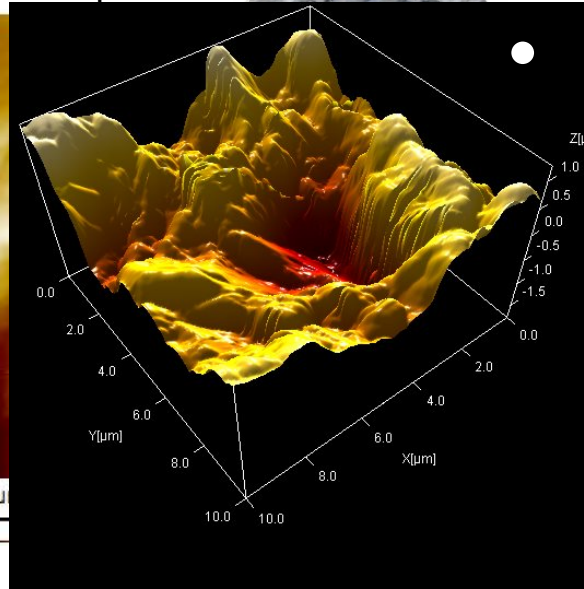
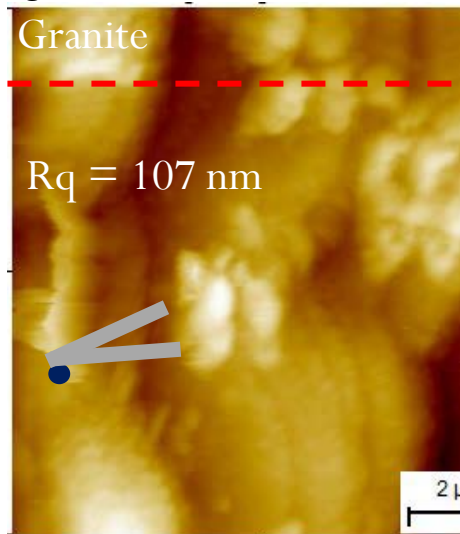
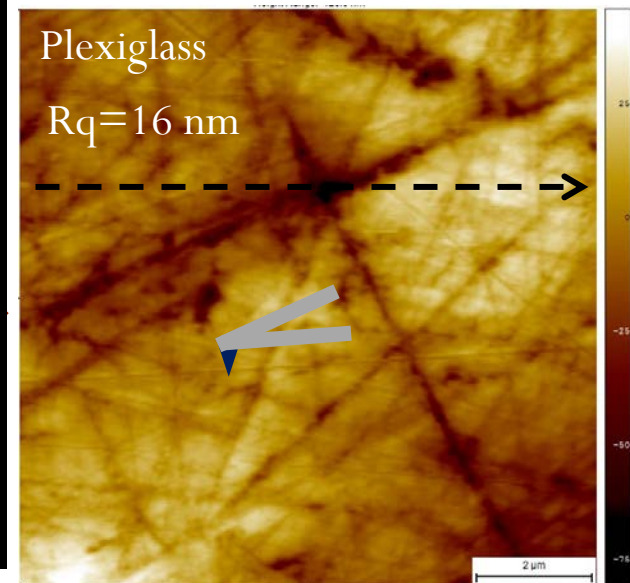
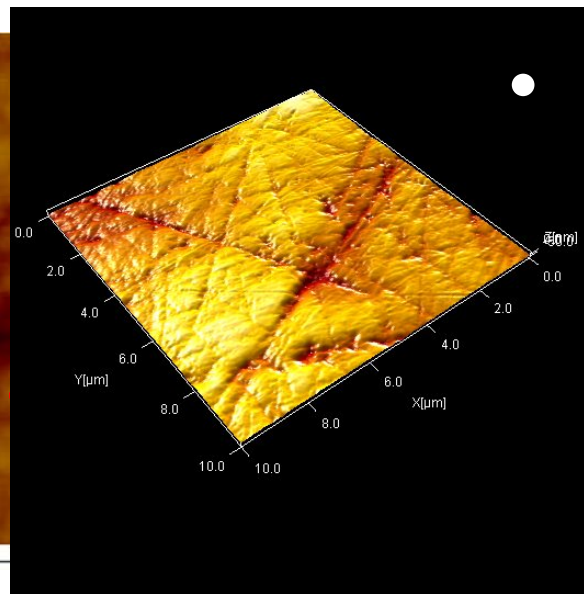
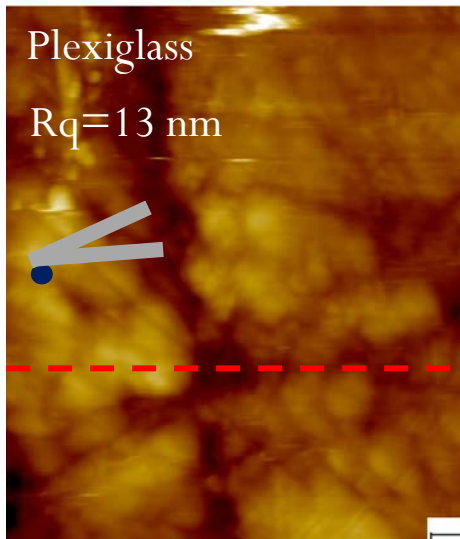
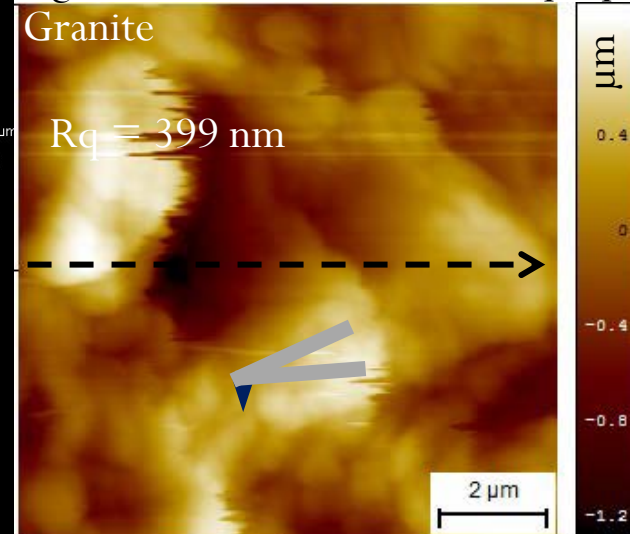
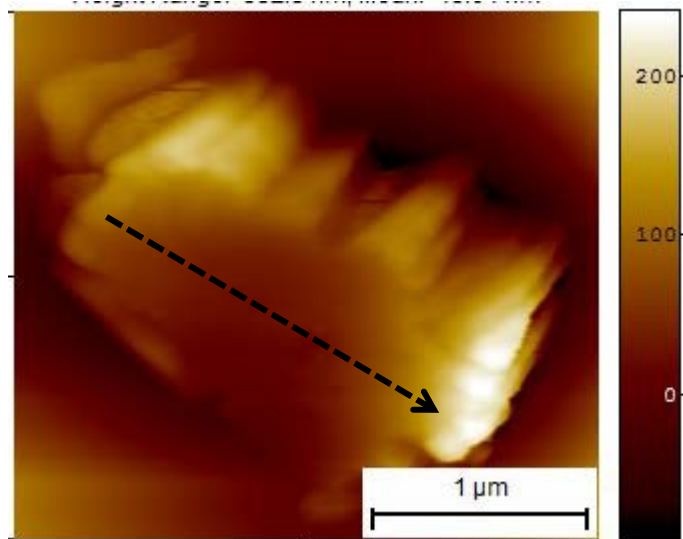


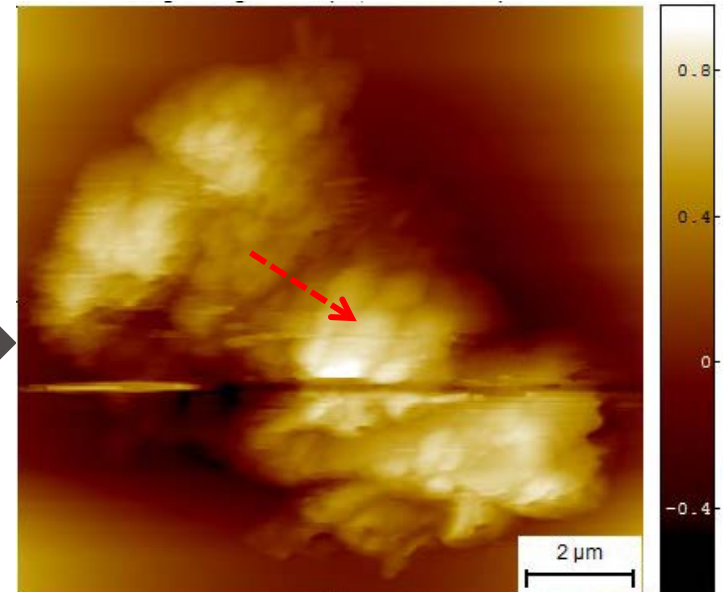
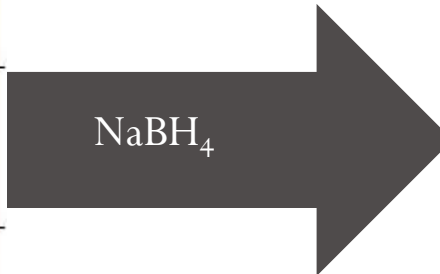
Image obtained with 24 nm sharp tip



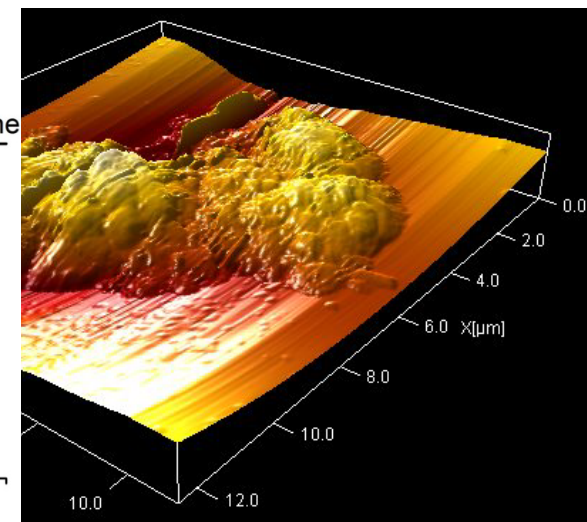
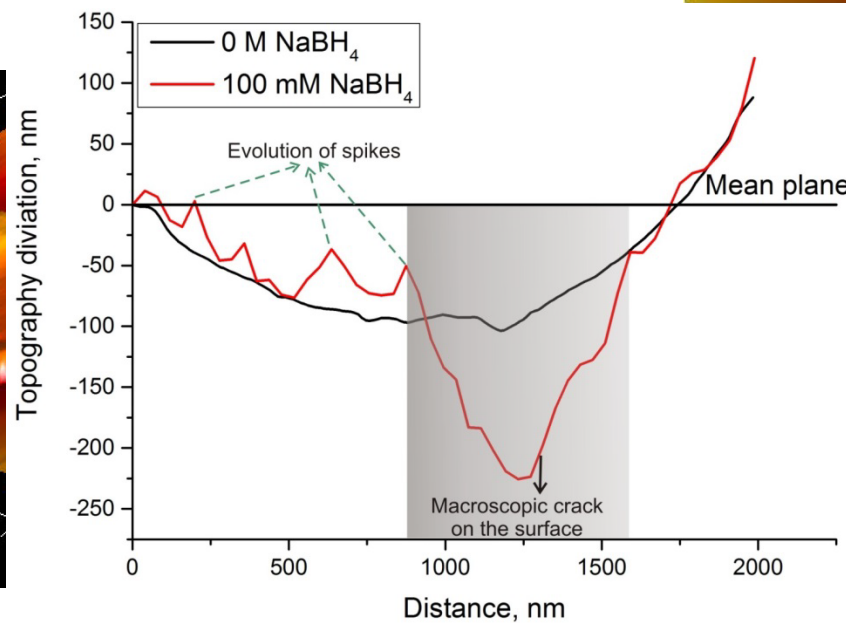
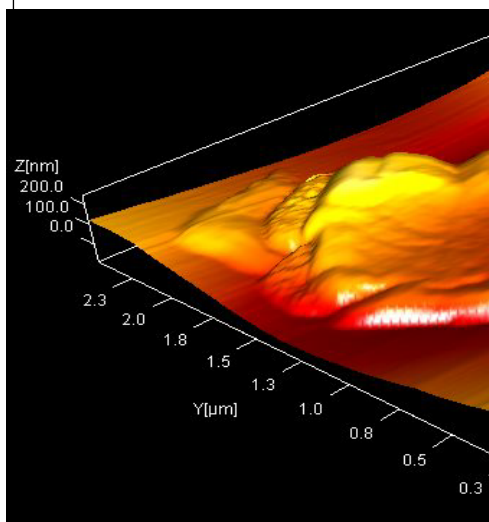
Magnetite surface topography change with NaBH_4



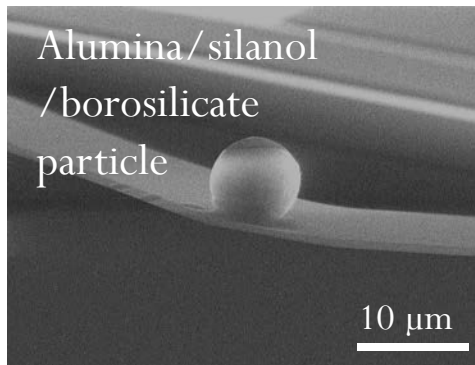
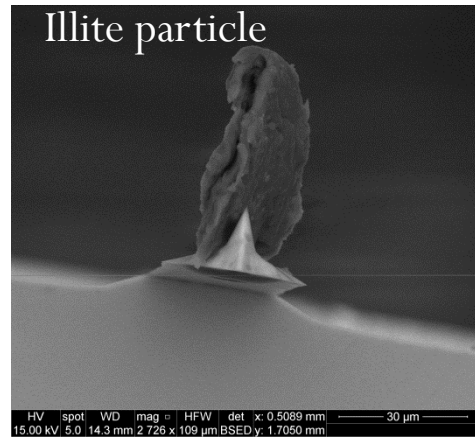
Magnetite in milliQ water



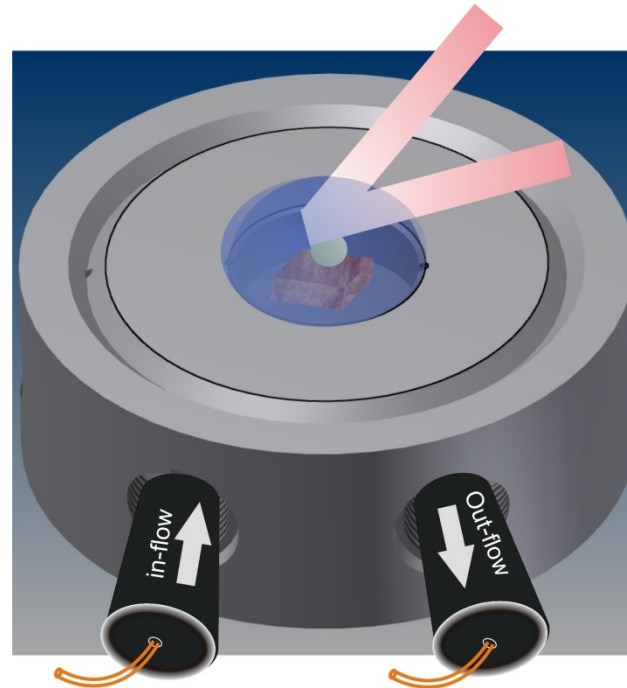
Magnetite in 100 mM NaBH_4



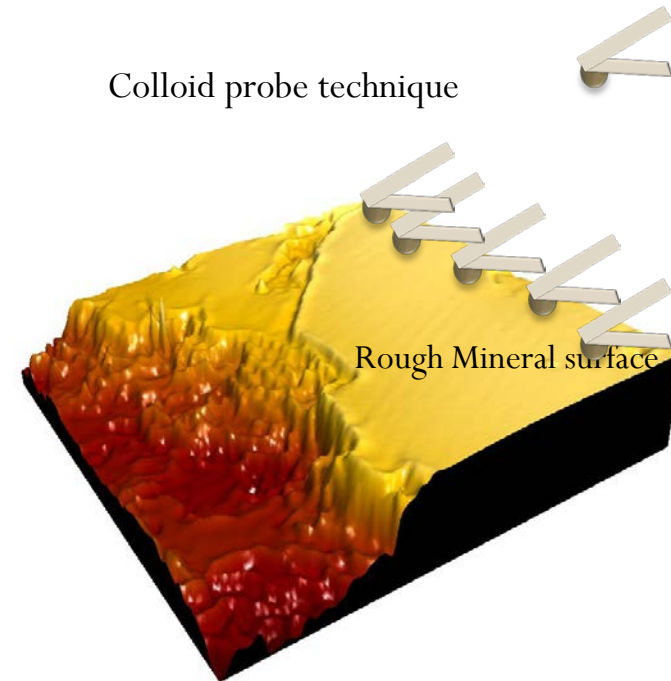
Force measurements



SEM of Al_2O_3 particle
attached to cantilever



Home made continuous flow through-fluid
cell for AFM experiments

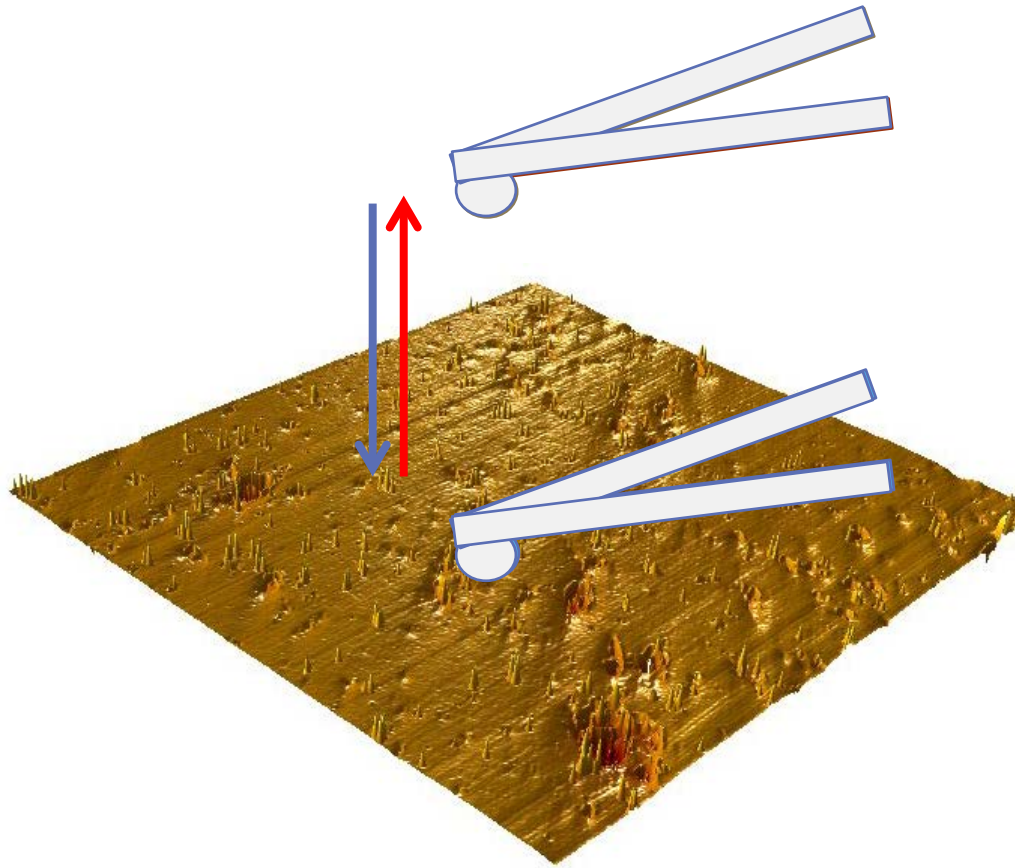


Parameters measured:

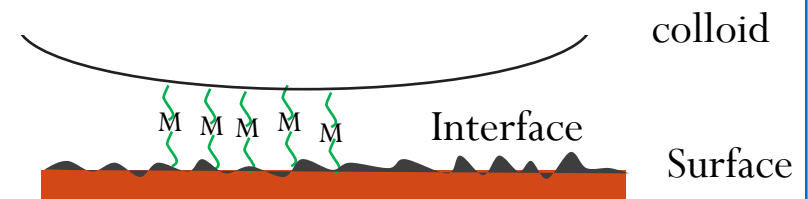
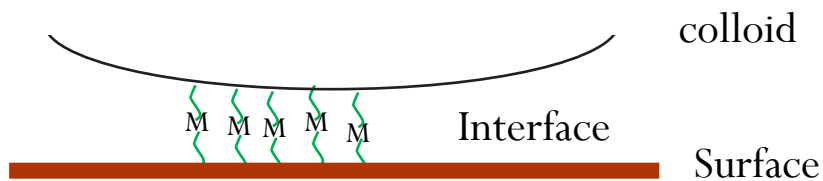
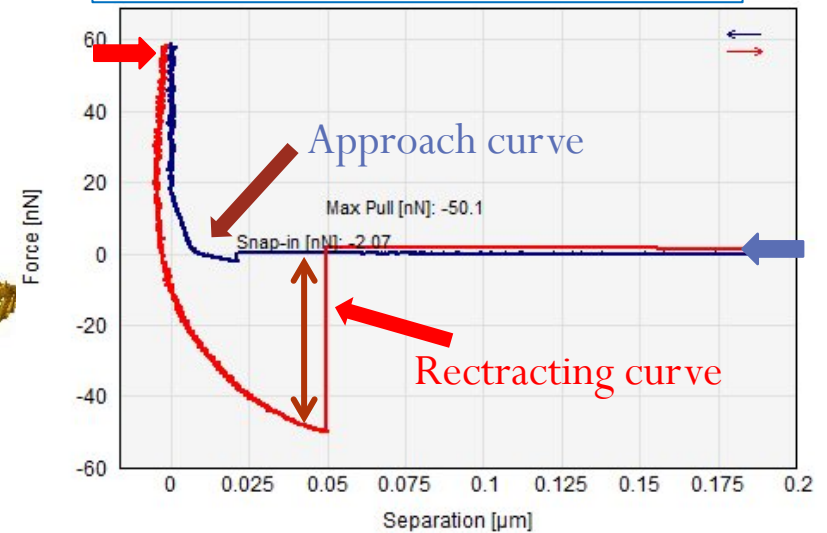
1. F_{Adhesion}
2. Snap-in force
3. Force-volume measurements

Applying colloid probe technique to measure the surface forces between
colloid particles (polystyrene, Al_2O_3) and rough minerals

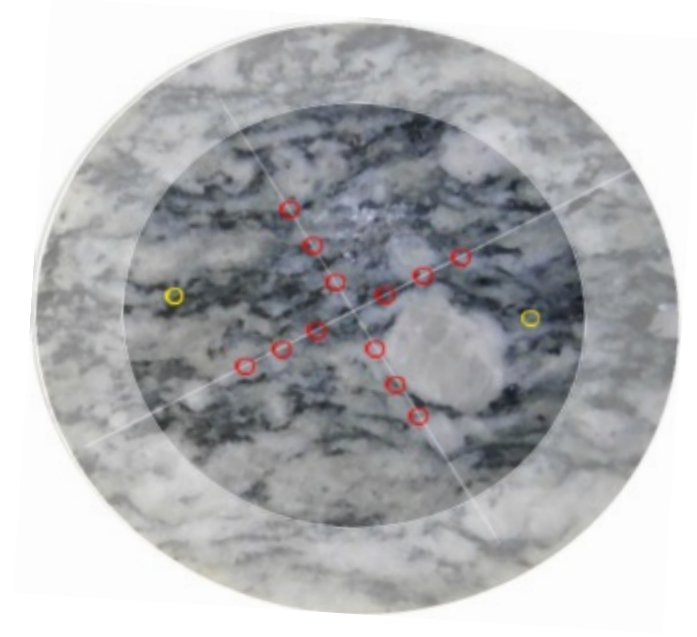
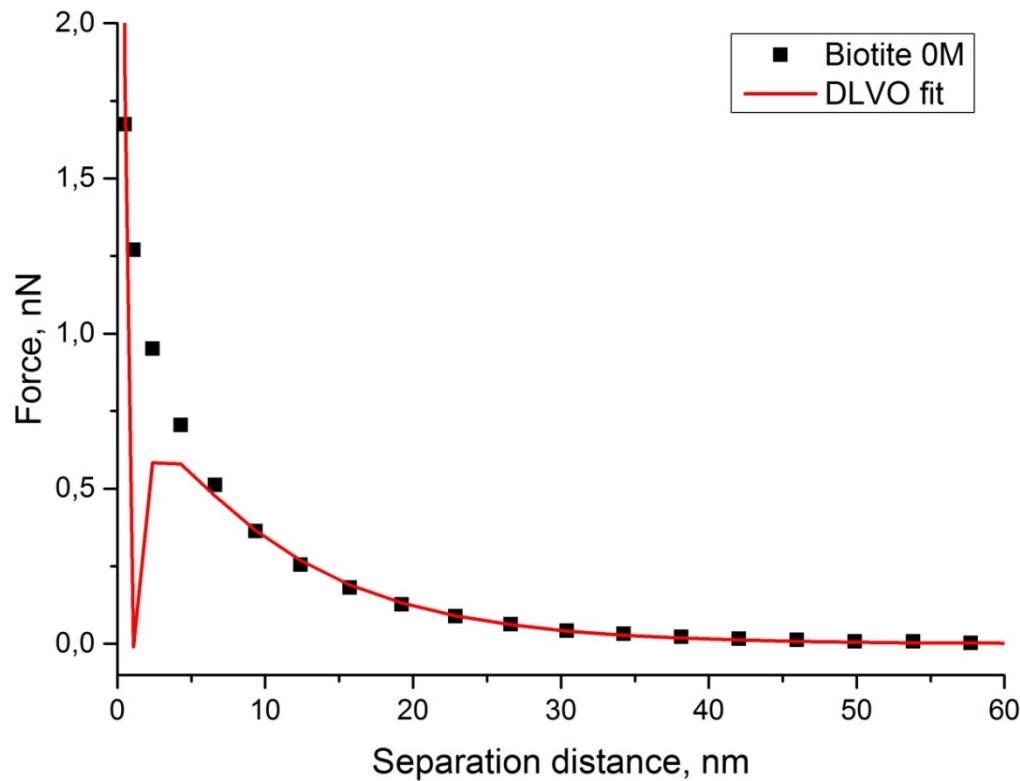
Colloid probe technique



Parameters obtained:
Surface potential
Young's modulus
Hamaker's constant
Dissipated Energy etc..



Fitting of AFM force curve with DLVO equation to obtain unknown potential of substrate

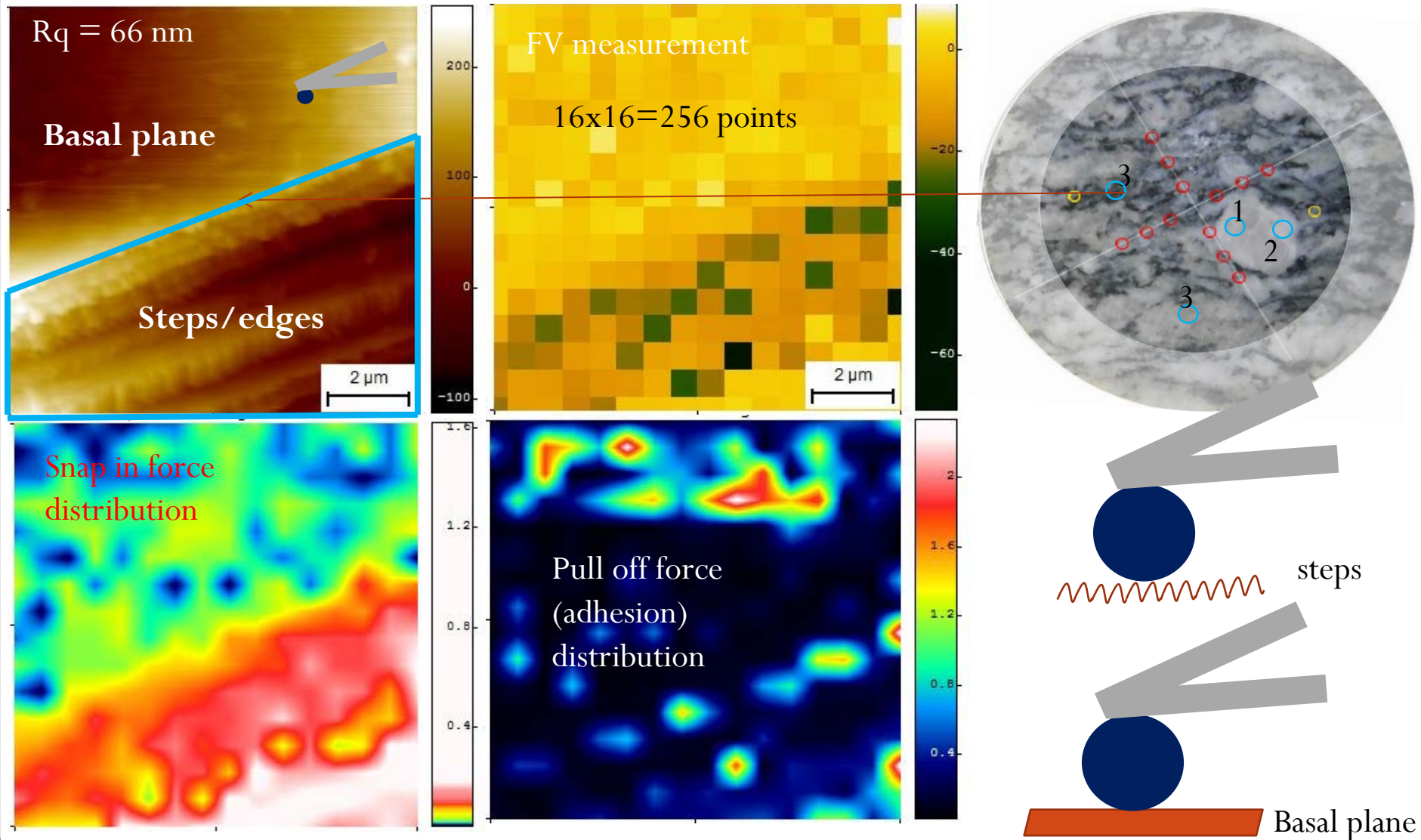


Zeta potential of colloid (measured using zeta sizer) = -32.4 mV

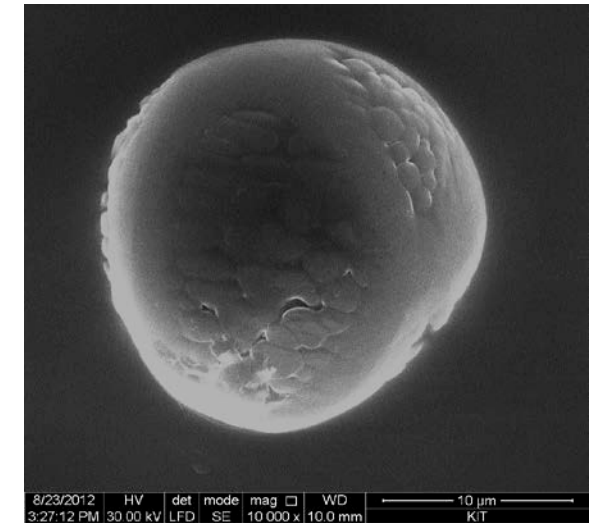
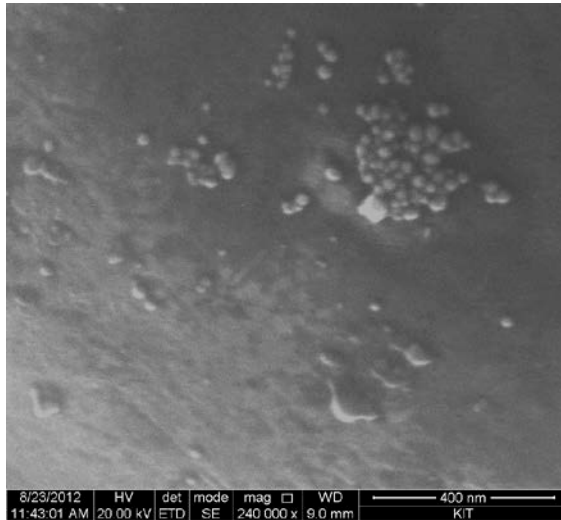
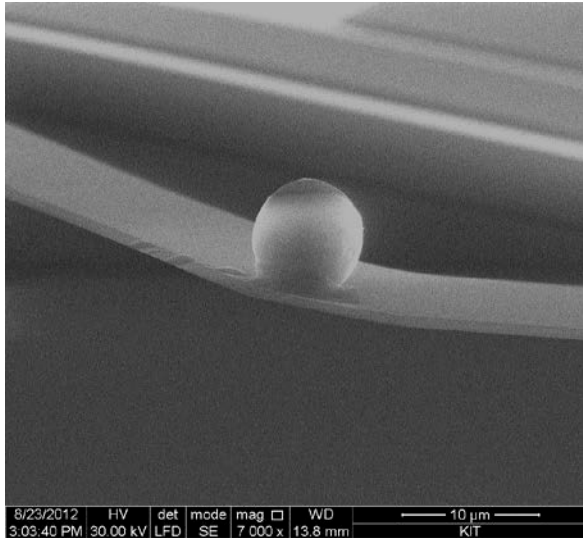
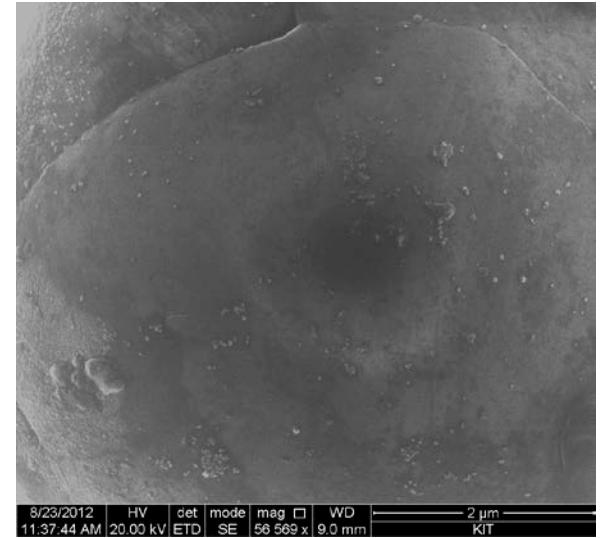
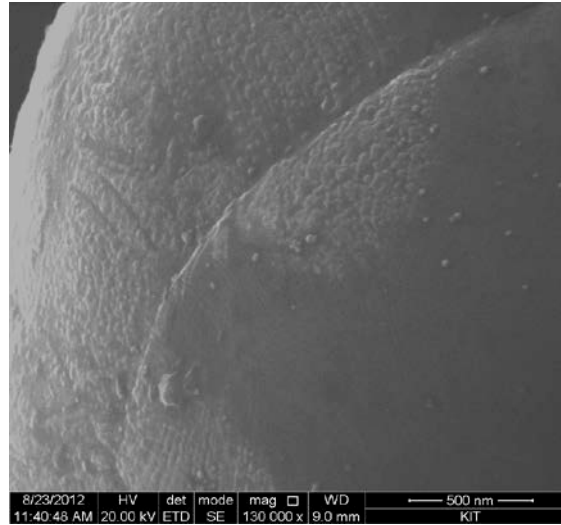
Zeta potential obtained after fitting the AFM approach force curve using DLVO equation = -74.2 mV

Force-volume measurements (Force+imaging)

Biotite/Dark minerals (mica) in 1 mM NaCl at pH 5

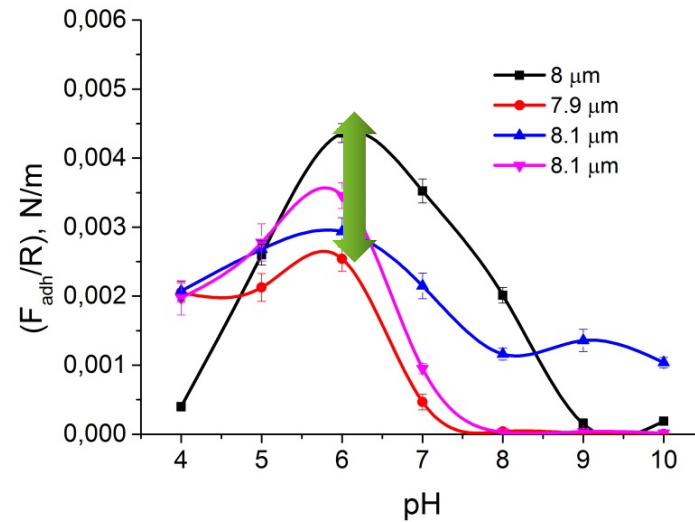


SEM of Al_2O_3 particles at high magnification

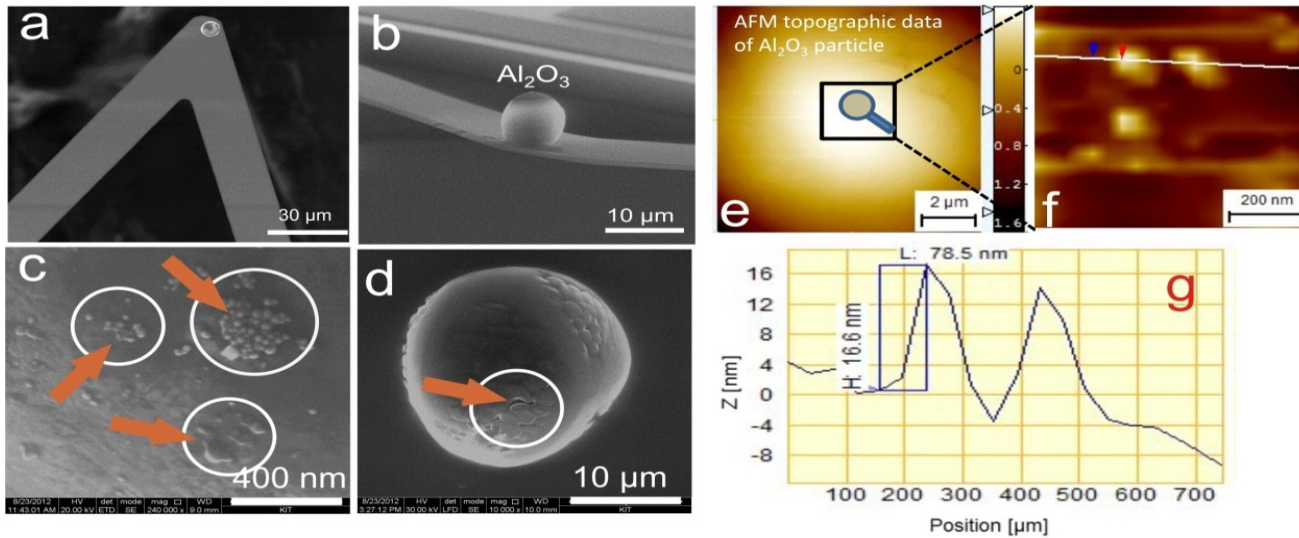


The SEM images clearly demonstrate: at higher magnification, there is a sub-micron scale roughness contribution from the colloids that could vary surface forces measured at the interface.

Different Al_2O_3 colloid probes Vs Biotite at varying pH



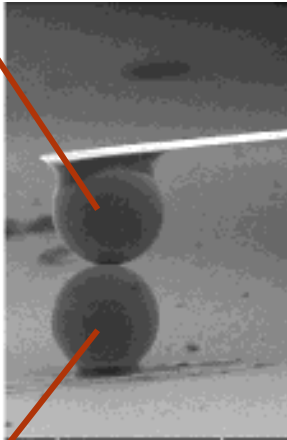
Adhesion forces
between alumina
particle and biotite



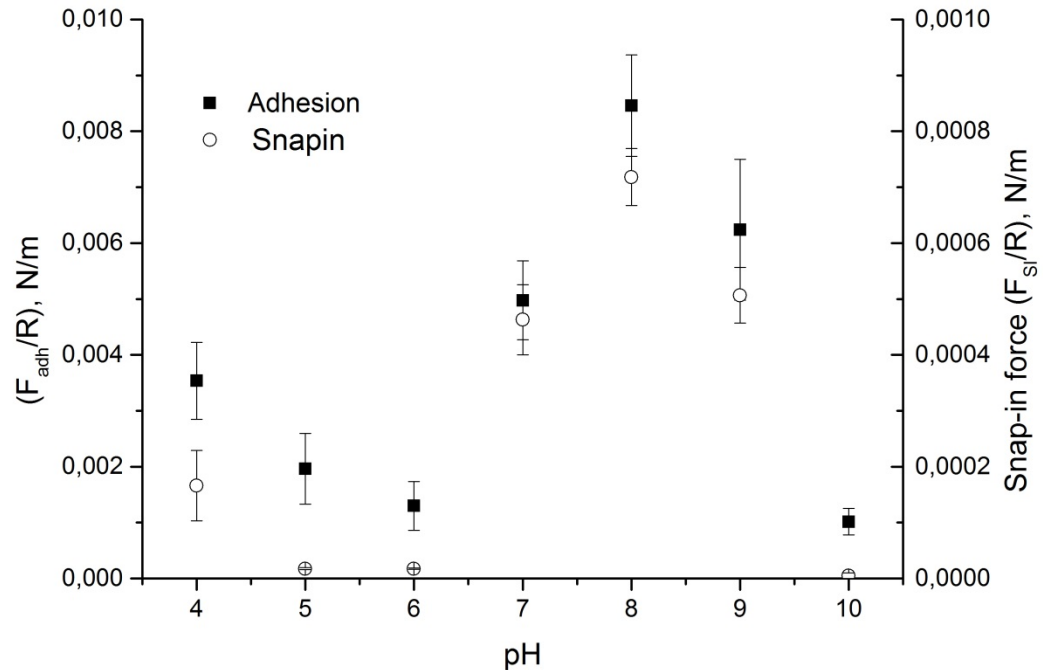
Roughness features on Al_2O_3 particle of <50 nm have pronounced effect on adhesion forces

Interaction between Al_2O_3 colloid probe and Al_2O_3 particle as a function of pH at 1 mM NaCl

Particle on cantilever



Particle on silicon wafer
or other immobile
substrate



The point of zero charge of Al_2O_3 particles is at pH 8

Sample preparation

- Clay colloid characterization:



0,01 g / 10 mL



Clay suspension

50 μ L

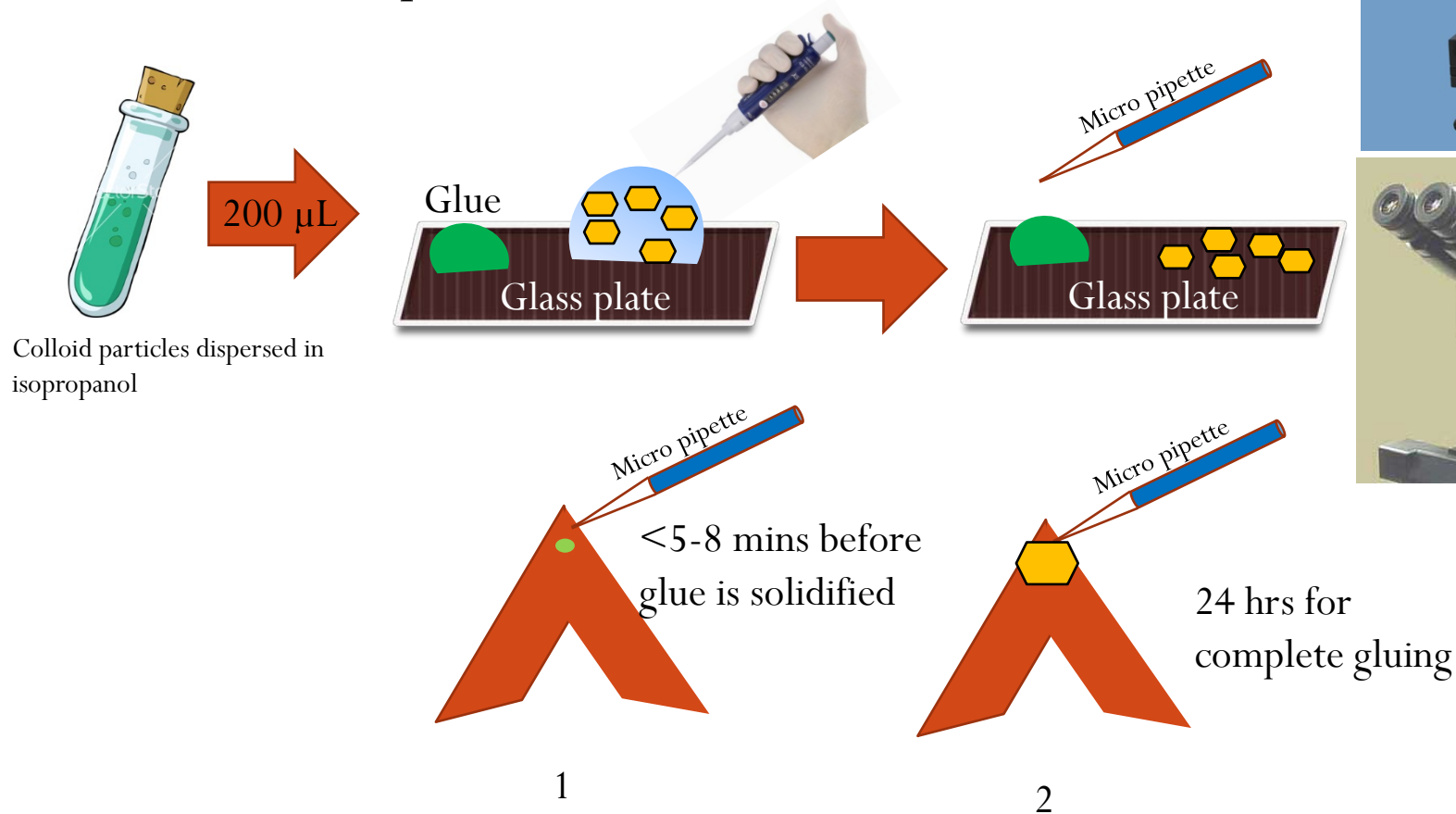


Allow it to dry for
measurements



Sample preparation

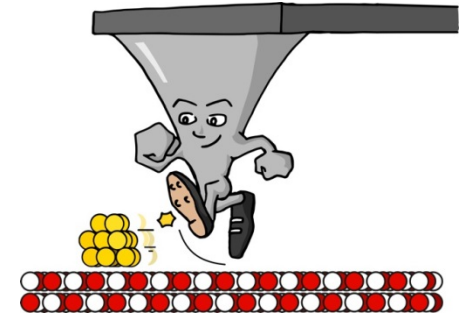
- Colloid probe for force measurements:



The samples are characterized in SEM before

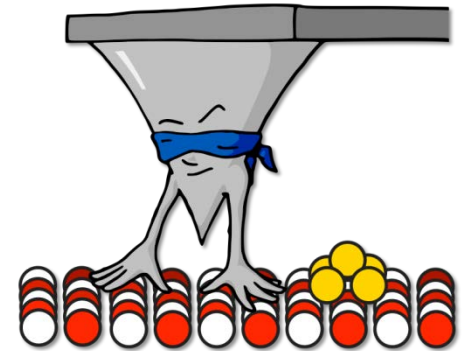
Conclusions....

1. AFM provides nano and submicron scale topographic information
2. It is versatile technique to measure the forces between surfaces
3. Finally...AFM is a blind technique



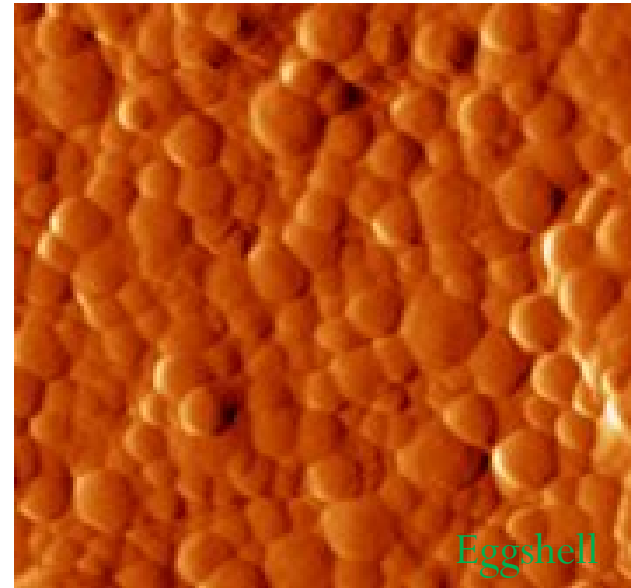
Limitations...

1. Sample should be steady, e.g. swelling clays: possibility of contamination to the tip
2. Substrates should not be too rough
3. The suspensions should not be too viscous
4. Particles attached should be $> 4 \mu\text{m}$
5. Etc....





Human hair



Eggshell